

Unveiling the Pathophysiological Landscape of COVID-19: Insights for Novel Therapeutic Strategies

Laiba Iqbal^{1*}, Dr. Naila Iqbal², and Muhmmad Milhan Afzal Khan³

¹Center of Excellence in Molecular Biology, University of the Punjab, Lahore, Pakistan.

(laiba.iqbal@cemb.edu.pk)

²College of Physicians and Surgeons, Pakistan. (e-mail: dr.nailaiqbal@gmail.com)

³Department of Computer Science, University of Agriculture, Faisalabad, Pakistan; (milhankhan@uaf.edu.pk)

*Corresponding author: Laiba Iqbal¹ (e-mail: laiba.iqbal@cemb.edu.pk).

Received: 20/01/2024, Revised: 15/04/2024, Accepted: 30/04/2024

Abstract- The ongoing COVID-19 epidemic has emphasised the critical need for a thorough understanding of the disease's pathophysiology environment. This article explores the complex relationship between SARS-CoV-2, the virus responsible for COVID-19, and the human body, offering light on the underlying mechanisms that determine illness susceptibility, development, and severity. The genetic, immunological, and physiological aspects that contribute to the different clinical presentations of COVID-19 using a multidisciplinary approach have been discussed. This article aims to provide significant insights that pave the way for potential therapeutic options addressing the core pathophysiological processes by deciphering these complex pathways. As the global community attempts to combat the epidemic, a fuller knowledge of COVID-19's molecular complexities holds the prospect of new interventions that can effectively lessen its effects. In addition, a case study featuring Quantitative Polymerase Chain Reaction (QPCR) amplification curves and Cycle Threshold (CT) values is shown to demonstrate the significance of making an accurate diagnosis. This diagnostic approach lets physicians to distinguish between positive and negative cases, assess viral load, and offer informed treatment recommendations. By combining QPCR results with clinical data, we can acquire a greater understanding of COVID-19's progression and influence the development of novel therapy methods. This research aims to aid the global effort to eradicate COVID-19 by elucidating its pathophysiological intricacies and employing sophisticated diagnostic techniques such as (QPCR).

Index Terms-- COVID-19, SARS-CoV-2, Pathophysiological, therapeutic treatment

I. INTRODUCTION

The COVID-19 pandemic has left an indelible effect on worldwide public health, with its origins traceable to December 2019 in Wuhan, China. As of August 16, 2023, the World Health Organization (WHO) releases staggering statistics: 769,806,130 confirmed illnesses and 6,955,497 deaths globally. As of the 19th of August, 2023, 13,499,865,692 vaccine doses had been provided to address this catastrophic health epidemic [1]. Figure 1 depicts the 7-day rolling average of daily hospitalizations of patients with COVID-19 diagnosis. At the centre of this global problem is SARS-CoV-2, a member of the Coronaviridae family responsible for COVID-19, a new Coronavirus disease that emerged in 2019. With a genomic size between 29.8 and 29.9 kilobytes [2], this virus resembles

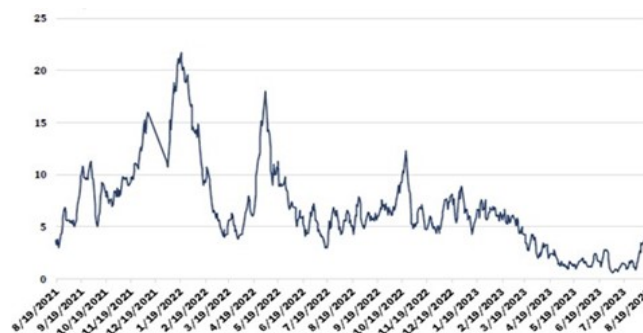


FIGURE 1. Daily Hospitalization with COVID-19 Diagnosis



This work is licensed under a Creative Commons Attribution –Strike Alike 4.0 International License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

its predecessor, SARS-CoV-1, which triggered a respiratory pandemic in the early 2000s [2].

Pathophysiology has a crucial link between the development of a disease and the manifestation of its symptoms. Consider, for example, the complex gear of the human respiratory system, a symphony of inhalation, oxygen exchange, and exhalation. Any discord in this symphony can result in a variety of lung illnesses, upsetting the delicate equilibrium of our respiratory physiology. Disease related to lungs, such as chronic obstructive pulmonary disease (COPD) and pneumonia, disrupt the respiratory harmony. In COPD, the airways become constricted, making breathing increasingly difficult, whereas pneumonia causes inflammation and fluid accumulation in the air sacs of the lungs, affecting their function. During the current pandemic, researchers have worked relentlessly to uncover the therapy choices and developing initiatives, from antiviral drugs to immunological regulators, in order to present a holistic assessment of the ongoing struggle against this worldwide health issue. To put an end to this unprecedented global health disaster, we need a comprehensive understanding of the pathophysiological complexities of COVID-19 to drive the development of innovative therapies and direct our concerted efforts.

The contribution of this paper are as follows:

- Analysed the complicated interplay between viral components, immune responses, and cellular pathways related to the pathophysiological environment of COVID-19.
- Evaluated therapy targets within the pathophysiology of COVID-19 including reducing viral replication, calming cytokine storms, and restoring endothelial function.
- Presented a QPCR case study on COVID-19 pathophysiology which provides a precise and accurate diagnostic method for identifying carriers and revealing viral load and infection state. These insights advance diagnostics and establish the framework for potential therapeutic approaches to reduce viral replication and its pathophysiological effects, as the article's goal.

The rest of the paper is organized as follows: Introduction section is followed by section II which discusses the structure and invasion system of COVID-19. Section III presents incubation time, typical symptoms, and transmission stages of COVID-19. In section IV, we analysed the body's reaction to COVID-19, paying special attention to the cytokine storm and the development of ARDS. It stresses the need of a well-functioning immune system and the risks associated with inefficient responses. Blood clot formation and the role of COVID-19 are studied in section V. This section also examines the role of endothelial dysfunction in this process. In section VI, we explore the role that age and gender play in determining one's susceptibility to COVID-19 and the severity of the condition. Detection methods for COVID-19 are covered in section VII. These include PCR testing, serological assays, and imaging procedures including chest X-rays and CT scans. Antiviral drugs, corticosteroids, monoclonal antibodies, and

pathophysiological secrets of COVID-19. Antiviral medications, immunomodulatory therapy, and vaccines owe a debt of gratitude to their accomplishments. Recent studies and published literature are analysed in this work to understand the pathophysiological landscape of COVID-19 and potential therapeutic strategies that may lead us to safer shores. To effectively navigate COVID-19, we must first understand its structure and invasion mechanisms, which will lead us through the phases and clinical stages of the disease and into the intricacies of the immune response, including the potentially lethal cytokine storms. We investigate the associations between age, gender, and genetic background and the severity and susceptibility to COVID-19. In addition, we investigate COVID-19 detection and monitoring options that make use of imaging and diagnostic methods. This essay finishes with a study of existing

convalescent plasma are just some of the treatment options for COVID-19 that are described in section VIII. Insight into potential therapy targets is provided in section IX on the basis of pathophysiological discoveries, including cytokine storms, endothelial dysfunction, and antiviral host factors. Section X presents a case study to analyse the significance of quantitative polymerase chain reaction (QPCR) in the diagnosis of COVID-19 and subsequent patient management. Finally, section XI concludes the study and presents future directions.

II. STRUCTURE AND INVASION SYSTEM

When a person coughs, sneezes or touches an object, the droplets of this virus is passed from that person to another. Figure 2[10] represents the structure of Coronavirus. It has the following structural components which are Spike protein (S), membrane protein (M), Envelope protein (E) and nucleocapsid protein (N) [2]. The receptors which provide way for different infections in human are ACE2, APN, Neu5 etc. [3]. Different

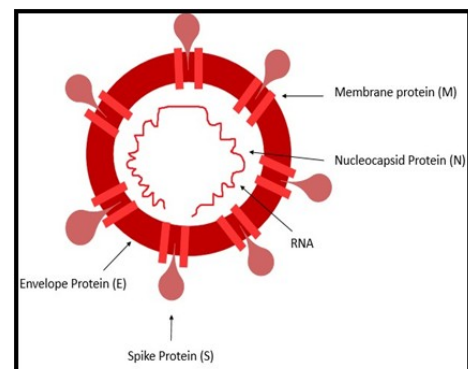


FIGURE 2. Structure of Coronavirus

experiments were carried out which revealed that SARS-COV- 2 uses ACE2 receptor (Angiotensin-converting Enzyme 2) for infecting humans [4]. The origin of infection is considered to be in the host bat (RaTG13) which is quite similar to the genomic sequence of SARS-COV-2 and the virus in bats. Coronavirus consists of positive

sense single stranded DNA. The most noticeable feature of this virus is club shaped spike proteins protruding from the exterior of the virus [5]. The S protein has two components: S1 which acts as a receptor binding domain, and S2, which forms the stalk that makes up the spike structure. The M protein is a protein in a virus that helps give it its shape. It is about 25-30kilo daltons [6], [7]. ACE2 receptor is found mostly in lungs, kidney and heart. S-proteins binds with ACE2 receptor which introduces COVID- 19's RNA into the cell. This binding also serves as a potential target for treatments and vaccinations [8], [9].

A. Clinical Symptoms and Disease Severity

The incubation period of COVID-19 is 5-6 days but can be up to 14 days. This period is also known as pre-symptomatic period. In this, the individual infected with the virus is contagious and has the ability to spread or transmit the virus to other individuals [11]. The common symptoms appearing in patients aged 40-60 years are fever, general body pain, dry cough, breathlessness and sometimes pneumonia in immuno compromised individuals or people battling an ongoing under- lying disease [12], [13]. Similar to other aerosol transmitted diseases, the transmission rate of COVID-19 is high. Gathering in a certain place can lead to transmission of this virus from one individual to another. Using different types of matricidal agents like ethanol (70%) can reduce the transmission rate [14].

III. PHASES/ CLINICAL STAGES OF COVID-19

There are three phases which indicate how COVID 19 progresses. Figure 3 indicates these phases which are incubation, symptomatic early and late pulmonary phase [15]. The late phase of incubation is the stage where patients infected with SARS-COV-2 are highly infectious. Human ACE2 receptor is expressed on ciliated epithelium of the nasopharynx and other nascent areas. ACE2 expression is much more in upper respiratory tract than lower [16]. After association with ACE-2, the engaged spike protein goes through a proteolytic cleavage which is infused by host membrane-anchored protein, the transmembrane protease serine 2 (TMPRSS2) [17]. There is spike mediated fusion at this stage, the virus then delivers the RNA genome and thus replication cycle starts [18]. Throughout the incubation and clinical phases, SARS-CoV-2 infects principally the ciliated epithelium of the nasopharynx and upper airways. Control of viral propagation is dependent on interactions between epithelial cells and immune cells, which are controlled by cytokine signalling and cell-to-cell contact. Viral infections are initially fought by innate immunity. Utilizing pattern recognition receptors, infected cells identify aberrant RNA structures upon virus entry (PRRs). These activate nuclear factor B (NF-B) and interferon regulator factors (IRFs). IRFs enhance antiviral defences by inducing

type I and type III interferons (IFN). The expression of pro-inflammatory cytokines and chemokines is triggered by NF-B, which recruits immune cells. However, SARS-CoV-2 suppresses the production of type I and type III IFN. Viral gene products such as NSP1, ORF6, ORF3B, and nucleocapsid (N) proteins inhibit signal transducer and activator of transcription1 (STAT1), hence diminishing interferon synthesis. The outcome of an infection is contingent upon viral load, replication rate, interferon response, and proinflammatory mediators [19], [20].

Patients with efficient interferon responses efficiently eradicate the infection. A function is played by a rapid reaction and robust innate immunity. A severe infection is the result of high viral replication and inadequate interferon responses. Its transmissibility is facilitated by the increased replication of the delta variant in the nasopharynx. Infected epithelial cells produce chemokines that attract immune cells such as macrophages, T cells, and mast cells [21], [22]. In severe situations, the virus may enter the lungs and infect type II pneumocytes and alveolar macrophages. This prepares for the pulmonary phase. Blood monocytes are drawn to infected lung tissue by a combination of a poor IFN response and increased chemokines. Macrophages express the enzymes necessary for viral fusion, ACE-2 receptor, furin, and TMPRSS2. Macrophages may function as Trojan horses, allowing viruses to anchor in the lungs [19].

Variability in ACE-2 expression among macrophages has the potential to affect the severity of an infection. Lower airway macrophages release pro-inflammatory cytokines and chemokines, which contribute to lung inflammation by recruiting additional immune cells. Activated platelets interact with monocytes to stimulate their recruitment. This chain of events contributes to lung inflammation [23]. In conclusion, SARS-interaction CoV-2's with the respiratory epithelium and the dynamics of the immune response play a critical role in determining the course of COVID-19 infection. Virus evasion of the interferon response, in conjunction with increased inflammation, leads to disease severity.

IV. IMMUNE RESPONSE AND CYTOKINE STORM

A cytokine storm often manifests as a flu-like symptom, which can develop more complicated as a result of injury to many organs. In extreme circumstances, a high fever is a common symptom, especially severe. Common symptoms include weariness, headache, joint discomfort, diarrhoea, lymph node enlargement, liver or spleen enlargement, altered feeling, and skin rashes. Frequently, fast respiration and low blood oxygen levels occur, which may lead to acute respiratory distress syndrome (ARDS). In severe situations, kidney disease, liver impairment, and stress-related cardiac problems may also occur [24].

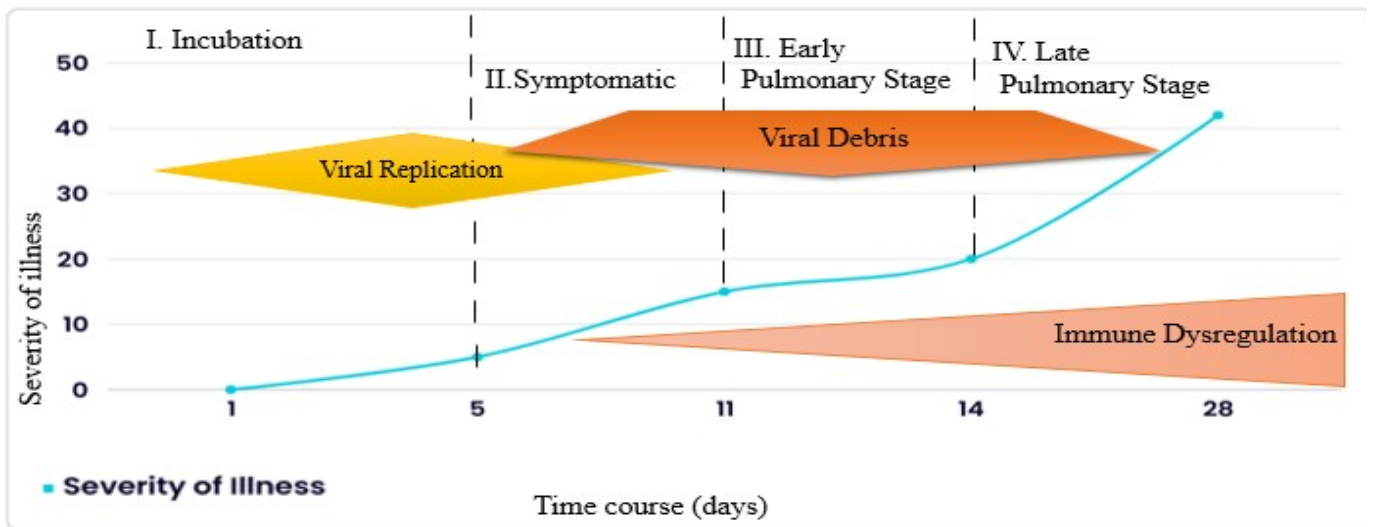


FIGURE. 3. Phases of COVID-19

The complications of a cytokine storm include capillary leak syndrome, which causes extensive edoema, and neurological complications such as encephalopathy. Blood clotting and bleeding irregularities develop, with the possibility of de-velopment to disseminated intravascular coagulopathy (DIC). Various blood cell count abnormalities and elevated levels of indicators such as C-reactive protein (CRP), ferritin, triglyc- erides, and D-dimer are detected by laboratory analysis [33]. Essentially, a cytokine storm begins with flu-like symptoms but can progress to serious organ damage and catastrophic problems involving several body systems. This may result in a range of clinical symptoms and test abnormalities.

A. Cytokine Storm and COVID-19

SARS-CoV-2 is a member of the family Coronaviridae, which includes enveloped, single-stranded, positive-sense ribonucleic acid (RNA) viruses that infect humans and other mammals [25]. It is airborne and mostly affects the respiratory system. Prior research showed that both severe acute respi- ratory syndrome coronavirus 1 (SARS-CoV-1) and MiddleEast respiratory syndrome coronavirus (MERS) are capable of inducing cytokine storms [26]. Following the development of SARS-CoV-2, the hypothesis of a cytokine storm gained attention as a significant contributor to COVID-19. Table 1 indicates key cytokines and regulatory mediators as presented the existing studies [26]–[30]. Multiple studies have found that COVID-19 patients have elevated levels of inflammatory cytokines such as IL-1, IL-2, IL-6, IL-10, IFN-, TNF-, IFN—inducible protein 10 (IP-10), granulocyte macrophage-colony-stimulating factor (GM-CSF), and monocyte chemoattractant protein-1 (MCP-1) [27]. Both biopsies and autopsy [28] have demonstrated the presence of inflammatory infiltrates in several tissues of COVID-19 patients. While the majority of cases demonstrate a self-

limiting flu-like symptom, vulnerable individuals with infection in lung cells, particularly type II pneumocytes, might experience an extensive influx of neutrophils, macrophages, CD8+ and CD4+ T lymphocytes, as well as considerable cytokine production. This chain of events results in bilateral pneumonia, acute respiratory distress syndrome (ARDS), and multi-organ dysfunction [29].

Insufficient immune response may account for the appearance of cytokine storms during SARS-CoV-2 infection. Infected cells generate large quantities of chemokines that recruit neutrophils and macrophages [30]. Interferon synthesis, which is essential for a successful antiviral response, is impaired in infected cells, while neutrophils and macrophages are recruited by the chemokines. Researchers have shown that COVID-19 patients produce autoantibodies against immune-regulatory proteins, in particular anti-type I interferon antibodies, which are associated with severe illness and mortality [31]. As seen in macrophages [32], the innate immune system may have difficulty clearing infected cells during the first phase, allowing the virus to proliferate freely. A following period of immunological recovery could result in an excessive response if viral multiplication continues unchecked.

Also hypothesised as a contributing factor [33] is the renin-angiotensin-aldosterone system (RAAS). SARS-CoV-2 enters human cells largely via angiotensin-converting enzyme 2 (ACE2), a member of the RAAS. By attaching to the AT1 receptor, ACE2 normally counteracts the pro-inflammatory, prothrombotic, and profibrotic actions produced by angiotensin

II. Virus binding to ACE2 may result in its downregulation within cells, potentially upsetting the equilibrium. This disturbance may result in reduced anti-inflammatory, antithrombotic, and anti-fibrotic effects, as well as enhanced angiotensin II-AT1 axis activity, possibly contributing to the cytokine storm and thrombo inflammatory condition observed in COVID-19.

TABLE I
KEY CYTOKINES AND REGULATORY MEDIATORS IN COVID-19

Cytokines/Inflammatory Mediators	Role in COVID-19	Possible Therapeutic Targets
IL-6	Extreme production associated with cytokines storm	Monoclonal antibodies, IL-6 suppressor
TNF- α	Pro-inflammatory cytokines	TNF suppressor
IFN- γ	Linked with immune dysregulation	Immune regulator
IL-1 β	Influences inflammation	IL-1 suppressor
IL-10	Inhibits immune response	Immune regulator
MCP-1	Attracts monocytes and macrophages	MCP-1 suppressor
GM-CSF	Excites immune cell response	Immune regulator

B. The progression of ARDS

Acute respiratory distress syndrome (ARDS) is a severe form of lung injury characterised by inflammation and increased permeability of the alveolar-capillary membrane, which leads to fluid accumulation in the lungs. The virus-induced inflammatory response can contribute to the development of ARDS in COVID-19 patients. The concentration of immune cells, including neutrophils and macrophages, in lung tissue can increase inflammation and result in the destruction of the lung's delicate structure [34], [35]. As lung tissue gets more inflammatory and fluid builds, the ability of the lungs to exchange oxygen and carbon dioxide is impaired. This causes significant respiratory distress, manifesting as shortness of breath, fast breathing, and low blood oxygen levels. In extreme circumstances, patients may need artificial ventilation to assist their breathing [36], [37].

V. LINK BETWEEN COVID-19 AND BLOOD CLOTTING DISORDER

Emerging data suggests that SARS-CoV-2 can cause individuals to develop a prothrombotic condition. It has been discovered that the virus directly infects endothelial cells, which line blood arteries. This endothelial infection can result in endothelial dysfunction, a condition in which the endothelium lacks the ability to maintain a healthy balance between vasodilation (increasing blood vessels) and vasoconstriction (narrowing blood vessels) (narrowing blood vessels). Endothelial dysfunction is a crucial element in the production of bloodclots [38].

- **Role of Endothelial Dysfunction:** Endothelial cells serve a crucial function in controlling blood flow, inhibiting platelet aggregation, and maintaining blood channel integrity. However, in COVID-19, the inflammation and immunological response caused by the virus might harm these cells, resulting in malfunction. When the endothelium is dysfunctional, it secretes fewer vasodilators (such as nitric oxide) and more vasoconstrictors, causing blood vessel constriction and contributing to clot formation [39].

Hypercoagulability associated with COVID-19: The endothelial dysfunction caused by SARS-CoV-2 infection can pave the way for hypercoagulability. Endothelial cells that have been compromised reveal tissue factor, a potent starter of the coagulation cascade. This increases the activation of platelets and the coagulation system. In addition to boosting the release of pro-inflammatory cytokines and

chemokines, the inflammation associated with COVID-19 exacerbates clot formation [40], [41].

VI. GENETIC VARIATIONS AND COVID-19 SUSCEPTIBILITY

Variations in genetics can influence the interaction between the SARS-CoV-2 virus and the host immune system. Variations in genes producing the ACE2 receptor, which the virus utilises to enter cells, and genes involved in immune responses can affect an individual's vulnerability to infection, for instance. Genetic variations in the human leukocyte antigen (HLA) system, which is responsible for delivering viral antigens to immune cells, can also alter the effectiveness of the immune response against the virus.

1) *Genetic Variations and COVID-19 Severity:* Variability in illness severity among COVID-19 patients is attributable in part to genetic variables. Genetic variations can affect the severity of the immune response, deciding whether a person mounts a protective response or develops an overactive inflammatory reaction, resulting in severe effects such as cytokine storms and acute respiratory distress syndrome (ARDS) [42]–[44].

2) *Age and Gender as Modifiers:* Important variables of COVID-19 severity are age and gender. Due to a reduced immune system and potential associated health issues, elderly adults are typically more prone to serious disease. Additionally, sex disparities in COVID-19 results have been noted, with males typically suffering from a more severe condition. It is hypothesised that genetic and hormonal factors contribute to these disparities [45], [46].

VII. METHODS OF DIAGNOSIS AND IMAGING FOR COVID-19 DETECTION

In contemporary medical practise, molecular diagnostics is a crucial frontier, with nucleic acid testing as its foundational technology. Among the many applications of nucleic acid testing, the accurate detection of the coronavirus has proven to be particularly effective. Nucleic acid amplification tests, such as Polymerase Chain Reaction (PCR) or isothermal nucleic acid amplification, antigen testing and other serological tests are the principal approaches for nucleic acid detection at present. The renowned sensitivity and specificity of PCR in detecting viruses requires heat cycling. Isothermal nucleic acid amplification, in contrast, provides rapid detection capabilities, operates at a constant temperature, and eliminates the requirement for a thermocycler [47]. Notably, as of March 24,

2020, the genome and proteome structures of the virus had been identified, however a thorough knowledge of the host's response to SARS-CoV-2 remained difficult [48]. Over one thousand COVID-19 sequences are currently available to the public. Figure 5 indicates the common diagnostic and imaging methods used for the detection of COVID-19.

1) *Polymerase Chain Reaction (PCR) Testing*: The gold standard for diagnosing COVID-19 is PCR testing. It recognises the genetic material of the virus in respiratory samples. It possesses a high degree of sensitivity and specificity, but requires specialised laboratories and a lengthy turnaround time [49]. DNA-amplification-based Polymerase Chain Reaction (PCR) tests are utilised for coronavirus identification. The reverse transcription of viral RNA into complementary DNA (cDNA) is a vital first step in these procedures, followed by PCR amplification for quantitative detection [50]. There are two ways to implement reverse transcription PCR (RT-PCR):

(a) *One-Step Assay*: This single-reaction technique includes cDNA synthesis and PCR amplification, decreasing mistakes but rendering it unsuitable for analysing many samples due to rapid RNA degradation. b) *Two-Step Assay*: This method, however more sensitive than the one-step approach, isolates reverse transcription from PCR amplification, enhancing sensitivity but increasing the danger of DNA contamination and necessitating more time [51]. Researchers have created highly sensitive and selective RT-PCR tests. Notably, the RdRp and E gene-based assays are frequently employed because of their high sensitivity, with a limit of detection (LOD) of 3.9 and 3.6 copies per 25 L reaction, respectively, whereas the N gene-based assay is less sensitive at 8.3 copies per 25 L reaction [52]. Real-time quantitative reverse transcription-PCR (rqRT-PCR) is preferred over traditional RT-PCR for coronavirus detection due to its superior specificity, quantitative analysis, and sensitivity. Numerous enhancements have been made to rqRT-PCR testing procedures. During amplification, the CDC uses one-step rqRT-PCR experiments that yield real-time fluorescence signals [53]. Researchers have created numerous rRT-PCR

assays targeting distinct genes for the detection of SARS-CoV-2. The RdRp/Hel assay is distinguished by its lower LOD of 11.2 RNA copies per reaction and the absence of cross-reactivity with other coronaviruses. In addition, innovative assays such as OSN-qRT-PCR and in-house real-time PCR techniques have shown great sensitivity [54]. In conclusion, PCR-based technologies, such as RT-PCR and rqRT-PCR, have become standard for sensitive and specific coronavirus identification, allowing a vast array of applications in the diagnosis of COVID-19. Researchers continue to improve the efficacy and dependability of these procedures.

2) *Antigen Testing*: Antigen testing detect viral proteins and deliver results quickly. They are less sensitive than PCR, but provide rapid findings, making them useful for preliminary

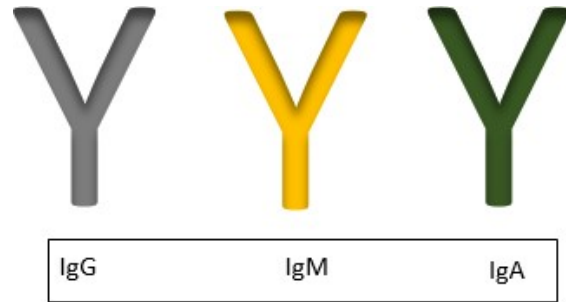


FIGURE 4. SARS COV-2 Antibodies

screening [55]. Rapid and extensive surveillance approaches are judged more effective than depending simply on nucleic acid amplification testing for suspected cases in controlling the spread of the virus [56]. Therefore, the development of reliable onsite detection technologies is urgently required.

By directly detecting viral antigens (Figure 1) or by recognising antibodies (Figure 4)[58], SARS-CoV-2 infection can be diagnosed. However, the possibility for cross-reactivity between SARS-CoV-2 antibodies and those developed in response to other coronaviruses poses a difficulty in generating appropriate serological testing. Lv et al. observed cross-reactivity between SARS-CoV-2 and SARS-CoV antibodies, which means that antibodies against SARS-CoV can be detected in plasma samples from SARS-CoV-2 patients [58]. This cross-reactivity is a substantial design challenge for serological diagnostics.

3) *Serological Assays*:: Serological tests detect SARS-CoV-2 antibodies in blood specimens. Due to the delay in antibody generation, they are less efficient for early detection but suggest past infection [49].

4) *X-rays*:: X-rays of the chest can indicate lung abnormalities such as infiltrates, opacities, and consolidation. Although they are not particular to COVID-19, they are useful for early evaluation [59].

5) *CT Scan*:: CT scans provide comprehensive images of lung tissue. They are sensitive in detecting lung abnormalities, aiding in the diagnosis of COVID-19 and determining the severity of the condition [60].

6) *Lung ultrasound*:: Lung ultrasound can highlight distinctive features such as B-lines and consolidations, aiding in early diagnosis and monitoring lung involvement [61].

VIII. EXISTING THERAPEUTIC METHODS

In the midst of the global struggle against the COVID-19 pandemic, researchers and medical experts have actively investigated numerous therapeutic techniques to minimise the impact of the SARS-CoV-2 virus on affected persons. These tactics include a variety of pharmaceutical interventions and treatments that try to target various components of the disease, such as reducing viral replication and regulating the immune system. In this section, we examine the current COVID-19 treatment options, highlighting their mechanisms of action and

potential benefits. From antiviral medications such as

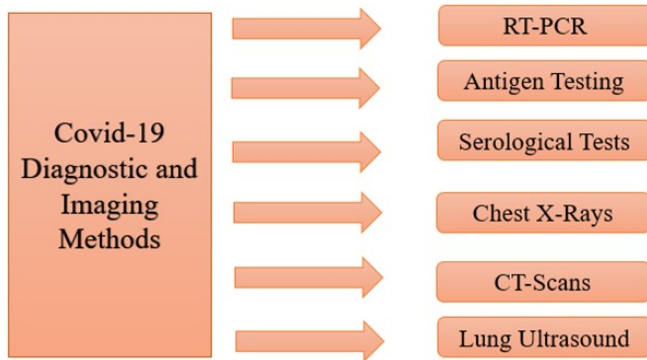


FIGURE 5. COVID-19 Diagnostics and Imaging Methods

remdesivir through the use of corticosteroids and monoclonal antibodies, we examine how these treatments have played a crucial role in illness management. In addition, we will investigate the viability of convalescent plasma obtained from recovered COVID-19 patients as a potential intervention in the ongoing war against the virus. Figure 6 indicates existing therapeutic methods for Covid-19.

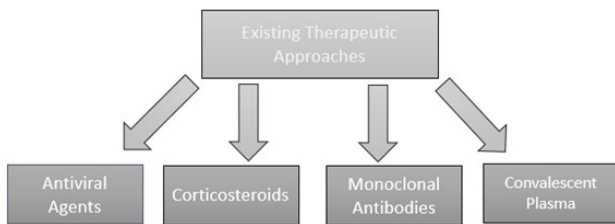


FIGURE 6. COVID-19 Diagnostics and Imaging Methods

1) *Antiviral Agents*: Drugs such as remdesivir are designed to prevent the SARS-CoV-2 virus' replication. Although they cannot eliminate the virus entirely, they can potentially lessen its spread and severity [62].

2) *Remdesivir*: Remdesivir, a C-adenosine nucleoside analog-derived monophosphoramidate prodrug, has emerged as a promising contender in the fight against COVID-19. It inhibits viral RNA replication by integrating itself into viral RNA chains, hence causing premature termination. Notably, remdesivir has significant anti-coronavirus action in the laboratory [63]. Previous research on coronaviruses, such as SARS-CoV and MERS-CoV, as well as bat CoV strains, indicated remdesivir's capacity to block reproduction in human airway epithelial cells. Additionally, it controls viral entry via the hACE2 receptor, indicating its ability to inhibit infection [64], [65]. Importantly, remdesivir acts in the early phases of infection, lowering RNA levels and virus titers dose- dependently [66]. In laboratory tests, remdesivir was effective against SARS-CoV-2, exhibiting an EC90 (90 percent effective concentration) value of 1.76 M in Vero E6 cells, which is comparable to its efficacy in non-human

primates [67]. Remdesivir's capacity to covalently bind to the RdRp gene, an essential component for SARS-CoV-2 replication, effectively halts the chain elongation process [68] is responsible for its efficacy. In addition, it possesses antiviral activity in human liver cancer cells, a cell line sensitive to SARS-CoV-2 [67]. With good outcomes, remdesivir has been delivered to COVID-19 patients for compassionate use outside of the laboratory. In these individuals, a 10-day course of the antiviral medication (beginning with 200 mg on day 1, followed by 100mg daily for 9 days) has demonstrated prospective clinical advantages [69]. Remdesivir's diverse antiviral activity makes it a promising candidate for further research in the fight against COVID-19.

3) *Corticosteroids*: such as dexamethasone, have been demonstrated to reduce an overactive immune response, particularly in severe cases, thereby minimising cytokine storms and their associated consequences [70].

4) *Monoclonal Antibodies*: Antibodies such as bamlanivimab and casirivimab/imdevimab can neutralize the virus, providing passive immunity and lowering viral load [71].

5) *Convalescent Plasma*: Plasma from recovered COVID-19 patients includes antibodies that can aid in the battle against the virus in infected individuals, however its effectiveness is still being studied [72].

IX PATHOPHYSIOLOGICAL FINDINGS INSIGHTS

Recent pathophysiological research on COVID-19 have Uncovered intriguing possibilities for therapeutic intervention. These investigations have extensively explored the complex mechanisms behind the infection and progression of the virus, suggesting possible therapeutic targets. By interpreting the molecular and cellular processes behind COVID-19, scientists have identified vulnerabilities that can be exploited to create innovative therapeutics. These results provide renewed hope to both patients and medical professionals, as they pave the way for more effective and individualized approaches to managing and combating the global health crisis. With continued research and worldwide cooperation, these new findings may eventually lead to game-changing advances in our fight against COVID-19. Table II presents summary of emerging therapeutic strategies as presented in existing studies [49], [55].

1) *Inhibition of Cytokine Storms*: Given the importance of cytokine storms in severe COVID-19 cases, targeted medicines to limit excessive inflammation, such as IL-6 inhibitors, could reduce the severity of the condition [73].

2) *Endothelial Dysfunction*: Since endothelial dysfunction leads to clotting problems, medications that target endothelial function may aid in the management of COVID-19-related coagulopathy [74].

3) *Antiviral Host Factors*: Comprehending the host factors necessary for viral replication could lead to the creation of

medications that inhibit these factors, thereby halting virus propagation.

IX. QPCR ANALYSIS FOR COVID-19 DETECTION: A CASE STUDY

In order to explain the complex pathophysiology of COVID-19 and pave the way for the development of novel therapeutic solutions, it is crucial to emphasize the necessity of precise

TABLE II
SUMMARY OF EMERGING THERAPEUTIC STRATEGIES

Therapeutic Approach	Mode of Action	Potential Benefits
Monoclonal Antibodies	Neutralize specific Cytokines	Reducing cytokine storm
Antiviral Agents	Suppress viral replication	Decrease in viral load
Immune Regulators	Regulation of immune response	Prevention of hyper inflammation
ACE Suppressors	Modulate renin-angiotensin system	Managing inflammation
RNA Vaccines	Eliciting immune response against virus	Immunization

and highly sensitive diagnostic approaches. These diagnostic methods serve as the basis for our understanding of the disease and play a crucial role in determining the development of novel treatments. These can provide the means to identify and analyse the virus’s activities within the human body, allowing healthcare practitioners and researchers to track its evolution and investigate the possibility of targeted interventions. The refinement and use of such accurate and sensitive diagnostic methods are essential to our scientific journey to understand COVID-19’s pathophysiological mechanisms and find novel treatment options. This case study examines the use of Quantitative Polymerase Chain Reaction (QPCR) for detecting the presence of SARS-CoV-2, the agent responsible for COVID-19, and demonstrates how these findings contribute to a greater knowledge of the disease.

A. Experimental Procedure

Using a specialist SARS-CoV-2 promoter kit or Meril COVID-19 one step PCR kit, BSL-3 containment-level laboratory conducted QPCR analysis. This diagnostic method targets particular genomic areas, such as the nucleocapsid (N) gene, the envelope protein (E) gene, and the ORF1ab gene. FAM, HEX and ROX are components of Meril COVID-19 pcr kits. FAM targets ORF1ab gene in Meril COVID-19 kit while it targets N gene if promotor kits are being used. Similarly, HEX targets N gene while in promotor kit, VIC dye is used which targets E gene. ROX in Meril COVID-19 kits target RNase P (IC) gene while ROX in promotor kit targets ORF1ab gene. Human RnaseP gene (IC) is the endogenous human positive control, it must be amplified in each sample. For promotorkit, for a patient’s sample to be positive, the CT value must be below 40 while for Meril kits, CT valye should be below 35. The QPCR analysis requires reverse transcription to convert viral RNA into complementary DNA (cDNA), followed by amplification with specified primers and fluorescence probes

tagged with different dyes. Monitoring fluorescence signals inreal time enables the detection of viral genetic material

B. qPCR graphs analysis

Establishing a key criterion is essential when analysing qPCR graphs, especially those from Meril COVID-19 kits. This important criterion involves ROX amplification, an assayreference signal. A strong ROX signal in every sample ismost important. ROX amplification ensures test a accuracy and reliability as an internal control. If ROX fails to amplify inany sample during qPCR, the test may be flawed. In such cases, discretion and diligence are needed. The patient’s test must be redone if ROX amplification is absent. The sample cannot be deemed positive or negative until ROX amplification is achieved. This thorough method ensures diagnostic validity and reduces the chance of incorrect or misleading outcomes. When using Meril COVID-19 kits, qPCR graph interpretation requires consistent ROX amplification. It ensures the diagnos-tic process’s precision and reliability, ensuring COVID-19 test results’ accuracy. Figure 7 indicates a highly positive case of COVID-19 of a patient. The value of ORF1ab gene is 22.65 and value of N gene is 23.08. This indicates that the viralload of patient is very high. Thus confirming that the patient is COVID positive.

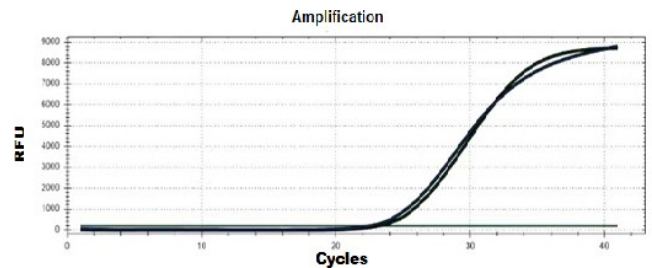


FIGURE 7. PCR result in which ROX was deselected, FAM and HEX channels were selected. Result of a sample indicated a sharp sigmoid curve indicating positive case

1) *Significance of the QPCR analysis:* QPCR analysis is a crucial diagnostic method for identifying COVID-19 individuals with precision and efficiency. The amplification curves and CT values provide information regarding viral load and infection state, hence facilitating patient care and epidemiological surveillance. By combining QPCR data with clinical data, researchers acquire a full knowledge of the course and severity of disease.

The use of QPCR in the case study corresponds with the article’s main objective of revealing the pathophysiological landscape of COVID-19. By identifying the presence of viral genetic material, QPCR illuminates the complex interplay between the virus and the molecular machinery of the host. These findings not only contribute to diagnostics, but also provide a platform for the development of novel therapeutic techniques aiming at reducing viral replication and its associated pathophysiological effects. As the goal of understanding the complexity of COVID-19 continues, QPCR is a crucial approach for revealing the disease’s molecular complexities,

hence influencing the development of innovative medicines and therapy modalities.

Figure 8 indicates a clear indication of the patient being Covid negative. The ORF1ab and N gene are targets of covid-19 but both came out negative. The curved lines below indicate noise.

C. Results and Discussion

Table III presents 96 samples, including positive and negative controls, on which qPCR was performed to assess the presence or absence of COVID-19. Table displays the Cycle Threshold (CT) values obtained for the samples under study. Sample 75 and Sample 49 had CT values of 16.00 and 24.64, respectively. These numbers indicate the amount of viral RNA present in the samples. Sample 75 had a lower CT value, indicating a larger viral load, whereas Sample 49 had a CT value that was moderately higher, indicating a lower viral load. Following standard laboratory procedures, these CT results were utilised to determine the presence (CT [threshold value]) or absence (CT [threshold value]) of COVID-19 in the suspect samples (CT [threshold value]). The QPCR study produced amplification curves, each of which indicated a distinct consequence. Positive controls displayed sigmoidal curves with distinct peaks, which indicated the presence of the virus. Negative controls, on the other hand, exhibited no sigmoidal curve, showing the absence of the virus. Observing the amplification curves and calculating the cycle threshold (CT) values were used to evaluate patient samples. CT values

represent the cycle at which detectable viral genetic material is first detected. A good result is indicated by a CT value lower than 38 together with a sigmoidal curve. The appearance of a sigmoidal curve that does not meet the CT value criteria necessitates confirmation testing.

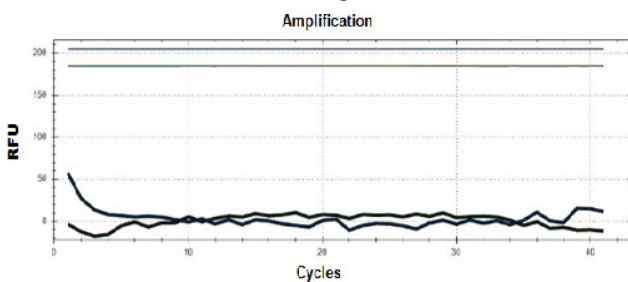


FIGURE 8. Patient sample from the same qPCR result indicating a negative result

X. CONCLUSION

In conclusion, the pathophysiology landscape of COVID-19 must be clarified for the development of successful therapeutic strategies. The wide spectrum of clinical consequences is driven by the intricate interactions between viral components, immunological responses, and cellular pathways. Although current therapy options are beneficial, they frequently focus on symptom management rather than addressing the underlying causes. Understanding the mechanism of the disease provides unique insights for treatment innovation. By targeting key pathways such as the

cytokine storm, endothelial dysfunction, and viral replication, a new generation of treatments can be developed to tackle COVID-19's underlying processes. As the global scientific community continues to progress, interdisciplinary collaborations will be essential for translating these insights into concrete and effective therapeutic strategies.

A. Future Directions/Perspectives

Future study is necessary to fill in the gaps in our understanding of COVID-19's pathogenesis. Investigating the genetic variants that affect disease susceptibility, elucidating the complex immune responses, and dissecting the mechanisms driving severe consequences will continue to be crucial. Integration of omics technologies, such as genomes and proteomics, can offer a comprehensive perspective on host-virus interactions. Utilizing cutting-edge computational approaches and artificial intelligence will aid in identifying prospective medication candidates. In order to translate pathophysiological findings into novel therapeutics, collaborations between researchers, doctors, and pharmaceutical corporations will be crucial. Ultimately, the development of creative therapeutic techniques guided by a thorough comprehension of COVID-19's biology gives promise in the fight against this unique worldwide health disaster.

TABLE III
PATIENTS SAMPLE CHECKED THROUGH QPCR TO FIND -
IVE AND +IVE COVID PATIENTS.

	1	2	3	4	5	6	7	8	9	10	11	12
A	26616	24	32	40	48	56	64	72	80	88	96	04
B	17	25	33	41	49 (24.64)	57	65	73	81	89	97	05
C	18	26	34	42	50	58	66	74	82	90	98	06
D	19	27	35	43	51	59	67	75 (16)	83	91	99	07
E	20	28	36	44	52	60	68	76	84	92	26700	08
F	21	29	37	45	53	61	69	77	85	93	01	09
G	22	30	38	46	54	62	70	78	86	94	02	+ve
H	23	31	39	47	55	63	71	79	87	95	03	-ve

ACKNOWLEDGMENT

The authors would like to thank University of Veterinary and Animal Sciences (UVAS) Lahore for providing an internship opportunity to work in BSL-3 lab.

REFERENCES

- [1] World Health Organization. Covid-19 data dashboard, 2023. Retrieved on 28th August, 2023, at 7:49 PM.
- [2] de Haan CA Rottier PJ. Bosch BJ, van der Zee R. The coronavirus spike protein is a class I virus fusion protein: structural and functional characterization of the fusion core complex. *Journal of virology*, 77(16):8801–8811, 2003.
- [3] Li F. Receptor recognition mechanisms of coronaviruses: a decade of structural studies. *Journal of virology*, 89(4):1954–1964, 2015.
- [4] Hong Z.-S. Tan Y.-Y. Chen S.-D. Jin H.-J. et al Guo Y.-R., Cao Q.-D. The origin, transmission and clinical therapies on coronavirus disease 2019 (covid-19) outbreak – an update on the status. *Military Med Res*, 7(4):11, 2020.
- [5] Vishwakarma K Nayak SK-Samantaray D Mohapatra S Agrahari R,

- Mohanty S. Update vision on covid-19: Structure, immune pathogenesis, treatment and safety assessment. *Sensors international*, 2(100073):8801–8811, 2021.
- [6] Jaimes JA and Whittaker GR. Feline coronavirus: Insights into viral pathogenesis based on the spike protein structure and function. *Virology*, 517:108–121, 2018.
- [7] Li F. Structure, function, and evolution of coronavirus spike proteins. *Annual review of virology*, 3(1):237–261, 2016.
- [8] Hooper NM Turner AJ, Hiscox JA. Ace2: from vasopeptidase to sars virus receptor. *Trends in pharmacological sciences*, 25(6):291–294, 2004.
- [9] Jr Siordia JA. Epidemiology and clinical features of covid-19: A review of current literature. *Journal of clinical virology : the official publication of the Pan American Society for Clinical Virology*, 127:104357, 2020.
- [10] Lingam G Seah I, Su X. Revisiting the dangers of the coronavirus in the ophthalmology practice. *Eye (London, England)*, 34(7):1155–1157, 2020.
- [11] World Health Organization. Coronavirus disease (covid-19) situation report, 2023. Accessed on 29th August 2023.
- [12] Leung GM Hedley AJ-Fraser C Riley S Abu-Raddad LJ Ho LM-Thach TQ Chau P Chan KP Lam TH Tse LY Tsang T Liu SH Kong JH Lau EM Ferguson NM Anderson RM Donnelly CA, Ghani AC. Epidemiological determinants of spread of causal agent of severe acute respiratory syndrome in hong kong. *Lancet (London, England)*, 361(9371):1761–1766, 2003.
- [13] Pinheiro LC Schenck EJ-Chen R Jabri A Satlin MJ Campion TR Jr-Nahid M Ringel JB Hoffman KL Alshak MN Li HA Wehmeyer GT Rajan M Reshetnyak E Hupert N Horn EM Martinez FJ Gulick RM Safford MM Goyal P, Choi JJ. Clinical characteristics of covid-19 in new york city. *The New England journal of medicine*, 382(24):2372–2374, 2020.
- [14] Henwood AF. Coronavirus disinfection in histopathology. *Journal of histotechnology*, 43(2):102–104, 2020.
- [15] Iglesias J Varon J-Cadegiani FA Marik PE Kory P, Meduri GU. Multi-modal hospital treatment protocol for covid-19 infection: Clinical and scientific rationale. *Journal of Clinical Medicine Research*, 14(2):53–79, 2022.
- [16] Edwards CE Martinez DR-Asakura T Dinnon KH 3rd Kato T Lee RE-Yount BL Mascenik TM Chen G Olivier KN Ghio A Tse LV Leist SR Gralinski LE Schäfer A Dang H Gilmore R Nakano S Baric RS Hou YJ, Okuda K. Sars-cov-2 reverse genetics reveals a variable infection gradient in the respiratory tract. *Cell*, 182(2):429–446.e14, 2020.
- [17] Müller MA Allen P-Soilleux E Pfeiffer S Steffen I Tsegaye TS He Y Gnirss K Niemeyer D Schneider H Drosten C Pöhlmann S Glowacka I, Bertram S. Evidence that tmprss2 activates the severe acute respiratory syndrome coronavirus spike protein for membrane fusion and reduces viral control by the humoral immune response. *Journal of virology*, 85(9):4122–4134, 2011.
- [18] Michel T McMurray JVV-Pfeiffer MA Solomon SD Vaduganathan M, Vardeny O. Renin-angiotensin-aldosterone system inhibitors in patients with covid-19. *The New England journal of medicine*, 382(17):1653–1659, 2020.
- [19] Karram T Heyman SN Abassi Z, Knaney Y. The lung macrophage in sars-cov-2 infection: A friend or a foe? *Frontiers in Immunology*, 11:1312, 2020.
- [20] Liu WC-Uhl S Hoagland D Møller R Jordan TX Oishi K Panis M Sachs D Wang TT Schwartz RE Lim JK Albrecht RA tenOever BR Blanco-Melo D, Nilsson-Payant BE. Imbalanced host response to sars-cov-2 drives development of covid-19. *Cell*, 181(5):1036–1045.e9, 2020.
- [21] Dreux M Nisole S Sa Ribero M, Jouvenet N. Interplay between sars-cov-2 and the type i interferon response. *PLoS Pathogens*, 16(7):e1008737, 2020.
- [22] Morrison TE Whitmore A Funkhouser W Ward JM Lamirande EW Roberts A Heise M Subbarao K Baric RS Frieman MB, Chen J. Sars-cov pathogenesis is regulated by a stat1 dependent but a type i, ii and iii interferon receptor independent mechanism. *PLoS Pathogens*, 6(4):e1000849, 2010.
- [23] Palhinha L Teixeira L Barreto EA Pão CRR Righy C Franco S Souza TML Kurtz P Bozza FA Bozza PT Hottz ED, Azevedo-Quintanilha IG. Platelet activation and platelet-monocyte aggregate formation triggers tissue factor expression in patients with severe covid-19. *Blood*, 136(11):1330–1341, 2020.
- [24] June CH Fajgenbaum DC. Cytokine storm. *The New England Journal of Medicine*, 383(23):2255–2273, 2020.
- [25] Ozaslan M Khailany RA, Safdar M. Genomic characterization of a novel sars-cov-2. *Gene reports*, 19, 2020.
- [26] Mastracchio A Frati P Fineschi V Maiese A, Bolino G. An immunohistochemical study of the diagnostic value of trem-1 as a marker for fatal sepsis cases. *Biotech Histochem*, 94(3):159–166, 2019.
- [27] Liu W Liu J Liu K Shang J Deng Y Wei S Chen L, Liu HG. Analysis of clinical features of 29 patients with 2019 novel coronavirus pneumonia. *Zhonghua Jie He He Hu Xi Za Zhi*, 43(0):E005, 2020.
- [28] Del Duca F Santoro P Manetti AC La Russa R Di Paolo M Turillazzi E Fineschi V Maiese A, Frati P. Myocardial pathology in covid-19-associated cardiac injury: A systematic review. *Diagnostics (Basel)*, 11(9):1647, 2021.
- [29] Del Duca F Santoro P Manetti AC La Russa R Di Paolo M Turillazzi E Fineschi V Maiese A, Frati P. Myocardial pathology in covid-19-associated cardiac injury: A systematic review. *Diagnostics (Basel)*, 11(9):1647, 2021.
- [30] Liu WC Uhl S Hoagland D Møller R Jordan TX Oishi K Panis M Sachs D Wang TT Schwartz RE Lim JK Albrecht RA tenOever BR Blanco-Melo D, Nilsson-Payant BE. Imbalanced host response to sars-cov-2 drives development of covid-19. *Cell*, 181(5):1036–1045.e9, 2020.
- [31] Le Voyer T Rosain J Philippot Q Manry J Michailidis E Hoffmann HH Eto S Garcia-Prat M et al. Bastard P, Gervais A. Autoantibodies neutralizing type i ifns are present in 4 uninfected individuals over 70 years old and account for 20 deaths. *Science Immunology*, 6:eabl4340, 2021.
- [32] Qu Y Zhu H Zhu Q Tong W Bao L Lv Q Cong J Li D-Deng W Yu P Song J Tong WM Liu J Liu Y Qin C Huang B Lv J, Wang Z. Distinct uptake, amplification, and release of sars-cov-2 by m1 and m2 alveolar macrophages. *Cell Discovery*, 7(1):24, 2021.
- [33] Timens W Hillebrands JL Navis GJ Gordijn SJ Bolling MC Dijkstra G Voors AA Osterhaus AD-et al. Bourgonje AR, Abdulle AE. Angiotensin-converting enzyme 2 (ace2), sars-cov-2 and the pathophysiology of coronavirus disease 2019 (covid-19). *Journal of Pathology*, 251:228–248, 2020.
- [34] Li X et al. Huang C, Wang Y. Clinical features of patients infected with 2019 novel coronavirus in wuhan, china. *The Lancet*, 395(10223):497–506, 2020.
- [35] Du R et al. Zhou F, Yu T. Clinical course and risk factors for mortality of adult inpatients with covid-19 in wuhan, china: a retrospective cohort study. *The Lancet*, 395(10229):1054–1062, 2020.
- [36] Brown M Sanchez E Tattersall RS Manson JJ; HLH Across Speciality Collaboration UK Mehta P, McAuley DF. Covid-19: consider cytokine storm syndromes and immunosuppression. *The Lancet*, 395(10229):1033–1034, 2020.
- [37] Calfee CS Sinha P, Matthay MA. Is a “cytokine storm” relevant to covid-19? *JAMA Internal Medicine*, 180(9):1152–1154, 2020.
- [38] van der Meer NJM et al. Klok FA, Kruij M. Incidence of thrombotic complications in critically ill icu patients with covid-19. *Thrombosis Research*, 191:145–147, 2020.
- [39] Steiger P et al. Varga Z, Flammer AJ. Endothelial cell infection and endotheliitis in covid-19. *The Lancet*, 395(10234):1417–1418, 2020.
- [40] Jimenez D et al. Bikdeli B, Madhavan MV. Covid-19 and thrombotic thromboembolic disease: Implications for prevention, antithrombotic therapy, and follow-up: Jacc state-of-the-art review. *Journal of the American College of Cardiology*, 75(23):2950–2973, 2020.
- [41] Warkentin TE Thachil J van der Poll T Levi M Iba T, Levy JH. Diagnosis and management of sepsis-induced coagulopathy and disseminated intravascular coagulation. *Journal of Thrombosis and Haemostasis*, 17(11):1989–1994, 2019.
- [42] Bujanda L et al. Ellinghaus D, Degenhardt F. Genomewide association study of severe covid-19 with respiratory failure. *New England Journal of Medicine*, 383(16):1522–1534, 2020.
- [43] Kumar A et al. Pujhari SK, Rathore DK. Genetic variation in immune system genes and their significance in covid-19. *Preprints*, 2021.
- [44] Green CA Hardwick HE Pius R Norman L Holden KA Read JM Dondelinger F Carson G-Merson L Lee J Plotkin D Sigfrid L Halpin S Jackson C Gamble C Horby PW Nguyen-Van-Tam JS Ho A Docherty AB, Harrison EM and ISARIC4C investigators. Features of 20,133 uk patients in hospital with covid-19 using the isaric who clinical

characterisation protocol: prospective observational cohort study. *BMJ (Clinical Research Ed.)*, 369:m1985, 2020.

- [45] He W Wu F Liu XF Han DM Liu S Yang JK Jin JM, Bai P. Gender differences in patients with covid-19: Focus on severity and mortality. *Frontiers in Public Health*, 8:152, 2020.
- [46] Morgan R Klein SL. The impact of sex and gender on immunotherapy outcomes. *Biology of Sex Differences*, 11(1):24, 2020.
- [47] M. Shen, Y. Zhou, J. Ye, A. A. Abdullah Al-Maskri, Y. Kang, S. Zeng, and S. Cai. Recent advances and perspectives of nucleic acid detection for coronavirus. *J Pharm Anal*, 10:97–101, 2020.
- [48] B. Udugama, P. Kadhiresan, H. N. Kozlowski, A. Malekjahani, M. Osborne, V. Y. C. Li, H. Chen, S. Mubareka, J. B. Gubbay, and W. C. W. Chan. Diagnosing covid-19: The disease and tools for detection. *ACS Nano*, 14:3822–3835, 2020.
- [49] Lippi G and Plebani M. The critical role of laboratory medicine during coronavirus disease 2019 (covid-19) and other viral outbreaks. *Clinical Chemistry and Laboratory Medicine*, 58(7):1063–1069, 2020.
- [50] D. Adachi, G. Johnson, R. Draker, M. Ayers, T. Mazzulli, P. J. Talbot, and R. Tellier. Comprehensive detection and identification of human coronaviruses, including the sars-associated coronavirus, with a single rt-pcr assay. *Journal of virological methods*, 122:29–36, 2004.
- [51] M. L. Wong and J. F. Medrano. Real-time pcr for mrna quantitation. *BioTechniques*, 39:75–85, 2005.
- [52] V. M. Corman, O. Landt, M. Kaiser, R. Molenkamp, A. Meijer, D. K. Chu, T. Bleicker, S. Bru'nink, J. Schneider, M. L. Schmidt, D. G. Mulders, B. L. Haagmans, B. van der Veer, S. van den Brink, L. Wijsman, G. Goderski, J. L. Romette, J. Ellis, M. Zambon, M. Peiris, and C. Drosten. Detection of 2019 novel coronavirus (2019-ncov) by real-time rt-pcr. Euro surveillance : bulletin Europeen sur les maladies transmissibles = European communicable disease bulletin, 25:2000045, 2020.
- [53] Centers for Disease Control and Prevention. Research use only real-time-rt-pcr protocol for identification of 2019-ncov, 2020.
- [54] H. Rahman, I. Carter, K. Basile, L. Donovan, S. Kumar, T. Tran, D. Ko, S. Alderson, T. Sivaruban, J. S. Eden, R. Rockett, M. V. O'Sullivan, V. Sintchenko, S. C. Chen, S. Maddocks, D. E. Dwyer, and J. Kok. Interpret with caution: An evaluation of the commercial ausdiagnostics versus in-house developed assays for the detection of sars-cov-2 virus. *Journal of clinical virology : the official publication of the Pan American Society for Clinical Virology*, 127:104374, 2020.
- [55] Mak GC, Cheng PK, Lau SS, Wong KK, Lau CS, Lam ET, Chan RC, and Tsang DN. Evaluation of rapid antigen test for detection of sars-cov-2 virus. *Journal of Clinical Virology*, 129:104500, 2020.
- [56] D. B. Larremore, B. Wilder, E. Lester, S. Shehata, J. M. Burke, J. A. Hay, M. Tambe, M. J. Mina, and R. Parker. Test sensitivity is secondary to frequency and turnaround time for covid-19 screening. *Science advances*, 7(1), 2021.
- [57] Guo Rong, Yulong Zheng, Yunfei Chen, Yiting Zhang, Peng Zhu, and Mohamad Sawan. Covid-19 diagnostic methods and detection techniques. *Encyclopedia of Sensors and Biosensors*, pages 17–32, 2023.
- [58] Hui Lv, Nicholas C. Wu, Owen T. Tsang, Meng Yuan, Ranawaka A. P. M. Perera, Wai Shing Leung, Ray T. Y. So, Jacky M. C. Chan, Gabriel K. Yip, Thomas S. H. Chik, Yong Wang, Cyrus Y. C. Choi, Yuping Lin, Wing W. Ng, Jingyi Zhao, Leo L. M. Poon, Joseph S. M. Peiris, Ian A. Wilson, and Chris K. P. Mok. Cross-reactive antibody response between SARS-CoV-2 and SARS-CoV infections. *Cell reports*, 31:107725, 2020.
- [59] Bai HX, Hsieh B, Xiong Z, Halsey K, Choi JW, Tran TML, Pan I, Shi LB, Wang DC, Mei J, Jiang XL, Zeng QH, Eggin TK, Hu PF, Agarwal S, Xie FF, Li S, Healey T, Atalay MK, and Liao WH. Performance of radiologists in differentiating covid-19 from non-covid-19 viral pneumonia at chest ct. *Radiology*, 296(2):E46–E54, 2020.
- [60] Patel R, Babady E, Theel ES, Storch GA, Pinsky BA, St George K, Smith TC, et al.. Report from the american society for microbiology covid-19 international summit, 23 march 2020: Value of diagnostic testing for sars-cov-2/covid-19. *mBio*, 11(2):e00722–20, 2020.
- [61] Soldati G, Smargiassi A, Inchingolo R, and et al. Proposal for international standardization of the use of lung ultrasound for patients with covid-19: A simple, quantitative, reproducible method. *Journal of Ultrasound in Medicine*, 39(7):1413–1419, 2020.
- [62] Dodd LE Mehta AK Zingman BS Kalil AC Hohmann E Chu HY Luetkemeyer A Kline S-Lopez de Castilla D Finberg RW Dierberg K Tapson V Hsieh L Patterson TF Paredes R Sweeney DA-Short WR Touloumi G Beigel JH, Tomashek KM and ACTT-1 Study Group Members. Remdesivir for the treatment of covid-19 - final report. *New England Journal of Medicine*, 383(19):1813–1826, 2020.
- [63] Travis K. Warren, Robert Jordan, Michael K. Lo, Adrian S. Ray, Richard L. Mackman, Veronica Soloveva, Dustin Siegel, Michel Perron, Roy Bannister, Hon C. Hui, Nate Larson, Robert Strickley, Jay Wells, Kelly S. Stuthman, Sean A. Van Tongeren, Nicole L. Garza, Ginger Donnelly, Amy C. Shurtleff, Cary J. Retterer, Dima Gharaibeh, and Sina Bavari. Therapeutic efficacy of the small molecule gs-5734 against ebola virus in rhesus monkeys. *Nature*, 531:381–385, 2016.
- [64] Timothy P. Sheahan, Amy C. Sims, Rachel L. Graham, Vineet D. Menachery, Lisa E. Gralinski, James B. Case, Sarah R. Leist, Krzysztof Pyrc, Joy Y. Feng, Iva Trantcheva, Roy Bannister, Young Park, Darius Babusis, Michael O. Clarke, Richard L. Mackman, Jenna E. Spahn, Christopher A. Palmiotti, Dustin Siegel, Adrian S. Ray, Tomas Cihlar, and Ralph S. ... Baric. Broad-spectrum antiviral gs-5734 inhibits both epidemic and zoonotic coronaviruses. *Science translational medicine*, 9:eaa13653, 2017.
- [65] Travis K. Warren, Jay Wells, Rekha G. Panchal, Kelly S. Stuthman, Nicole L. Garza, Sean A. Van Tongeren, Liby Dong, Cary J. Retterer, Brett P. Eaton, Gianluca Pegoraro, Shelley Honnold, Shanta Bantia, Pravin Kotian, Xin Chen, Brian R. Taubenheim, Lisa S. Welch, David M. Minning, Y. Sudhakar Babu, William P. Sheridan, and Sina Bavari. Protection against filovirus diseases by a novel broad-spectrum nucleoside analogue bcx4430. *Nature*, 508:402–405, 2014.
- [66] Maria L. Agostini, Erica L. Andres, Amy C. Sims, Rachel L. Graham, Tim P. Sheahan, Xiaotao Lu, Everett C. Smith, James B. Case, Joy Y. Feng, Robert Jordan, Adrian S. Ray, Tomas Cihlar, David Siegel, Richard L. Mackman, Michael O. Clarke, Ralph S. Baric, and Mark R. Denison. Coronavirus susceptibility to the antiviral remdesivir (gs-5734) is mediated by the viral polymerase and the proofreading exoribonuclease. *mBio*, 9:e00221–18, 2018.
- [67] Manli Wang, Ruiyuan Cao, Leike Zhang, Xinglou Yang, Jia Liu, Mingyue Xu, Zhengli Shi, Zhihong Hu, Wu Zhong, and Gengfu Xiao. Remdesivir and chloroquine effectively inhibit the recently emerged novel coronavirus (2019-ncov) in vitro. *Cell Research*, 30:269–271, 2020.
- [68] Wenjuan Yin, Chunhong Mao, Xiaoli Luan, Dingding Shen, Qiong Shen, Hui Su, Xiaoyu Wang, Fang Zhou, Wenjie Zhao, Min Gao, Sheng Chang, Yurong C. Xie, Gang Tian, Hualiang W. Jiang, Sheng C. Tao, Jianhua Shen, Yong Jiang, Hualiang Jiang, and H. Eric Xu. Structural basis for inhibition of the rna-dependent rna polymerase from sars-cov-2 by remdesivir. *Science (New York, N.Y.)*, 368:1499–1504, 2020.
- [69] Jonathan Grein, Norio Ohmagari, Dongyool Shin, George Diaz, Elisa Asperges, Antonella Castagna, Tom Feldt, Gary Green, Michael L. Green, Francois-Xavier Lescur, Emanuele Nicastrì, Robin Oda, Kenzo Yo, Eugenia Quiros-Roldan, Alexandra Studemeister, John Redinski, Syed Ahmed, Jacob Bernett, Deepa Chelliah, David Chen, and Timothy Flanigan. Compassionate use of remdesivir for patients with severe covid-19. *The New England journal of medicine*, 382:2327–2336, 2020.
- [70] Lim WS Emberson JR Mafham M Bell JL Linsell L Staplin N Brightling C Ustianowski A Elmahi E Prudon B Green C Felton T Chadwick D Rege K Fegan C Chappell LC Faust SN Jaki T RECOVERY Collaborative Group, Horby P and Landray MJ. Dexamethasone in hospitalized patients with covid-19. *New England Journal of Medicine*, 384(8):693–704, 2021.
- [71] Norton T Ali S Gao H Bhoire R Musser BJ Soo Y Rofail D Im J Perry C Pan C Hosain R Mahmood A Davis JD Turner KC Hooper AT Hamilton JD Baum A Kyratsous CA Weinreich DM, Sivapalasingam S and Trial Investigators. Regn-cov-2, a neutralizing antibody cocktail, in outpatients with covid-19. *New England Journal of Medicine*, 384(3):238–251, 2021.
- [72] Fairweather D Senefeld JW Bruno KA Klassen SA Carter RE Klompas AM Wiggins CC Shepherd JR Rea RF Whelan ER Clayburn AJ Spiegel MR Johnson PW Lesser ER Baker SE Larson KF Ripoll JG Andersen KJ Joyner MJ, Wright RS and Casadevall A. Early safety indicators of covid-19 convalescent plasma in 5000 patients. *The Journal of Clinical Investigation*, 130(9):4791–4797, 2020.
- [73] Moore JB and June CH. Cytokine release syndrome in severe covid-19. *Science (New York, N.Y.)*, 368(6490):473–474, 2020.
- [74] Breakey N Escher R and Lammle B. Severe covid-19 infection associated with endothelial activation. *Thrombosis Research*, 190:62, 2020.